



PALYNOLOGY OF BEACH SQUATTER-1,

OTWAY BASIN, AUSTRALIA

BY

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FOR BEACH PETROLEUM

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I SUMMARY

790m (cutts) : M. diversus Zone (possibly upper) : Early Eocene :
very nearshore marine : immature

810m (cutts) : L. balmei Zone : Paleocene : nearshore marine :
immature

820m (cutts) : mixed assemblage presumed L. balmei with minor
reworked T. longus/M. druggii elements, but could be Late
Cretaceous with extensive caving : presumed Paleocene with
Maastrichtian reworking : marginally marine : immature

840m (cutts) : T. longus/M. druggii Zones : Maastrichtian :
marginal marine : immature

1000m (cutts) : T. pachyexinus/N. aceras Zones : Santonian -
Campanian : nearshore to marginal marine : immature

1310m (cutts) : C. triplex Zone : Turonian : nearshore marine :
marginally mature for oil

1390m (cutts) - 1420m (cutts) : A. distocarinatus/P.
infusorioides Zones : Cenomanian : very nearshore marine :
marginally mature for oil

1500m (cutts) : P. pannosus Zone : late Albian : presumed
non-marine : marginally mature for oil

II INTRODUCTION

Ten cuttings samples were examined from Beach Squatter-1 for biostratigraphy and spore colour. No sidewall cores were available due to poor hole conditions. Yields were generally good. The samples are assigned to seven palynological zones on the basis of the supporting data presented here as Appendix I. The Cretaceous zonation used is basically that of Helby, Morgan and Partridge (1987), which draws on all previous work. The Tertiary zonation is that of Stover and Partridge (1973) and Stover and Evans (1973) as modified by Partridge (1976). Figure 1 shows the zonation framework.

Maturity data was generated on the Thermal Alteration Index (TAI) Scale of Staplin and plotted on Figure 2 as a Maturity Profile. The oil and gas windows on Figure 2 follow the general consensus of geochemical literature. The oil window corresponds to spore colours of light-mid brown (2.7) to dark brown (3.6) and would correspond to Vitrinite Reflectances of 0.6% to 1.3%. Geochemists, however, have not reached universal agreement on these values and argue variations based on kerogen type, basin type and basin history. The maturity interpretation is thus open to reinterpretation using the basic colour observations as raw data. However, the range of interpretation philosophies is not great, and would probably not move the oil window by more than 200 metres. Instrumental geochemistry offers quantitative and repeatable raw data.

AGE		SPORE - POLLEN ZONES	DINOFLAGELLATE ZONES	
Early Tertiary	Early Oligocene	<i>P. tuberculatus</i>		
	Late Eocene	upper <i>N. asperus</i>	<i>P. comatum</i>	
		middle <i>N. asperus</i>	<i>V. extensa</i>	
	Middle Eocene	lower <i>N. asperus</i>	<i>D. heterophlycta</i>	
			<i>W. echinosuturata</i>	
	Early Eocene		<i>P. asperopolus</i>	<i>W. edwardsii</i>
			upper <i>M. diversus</i>	<i>W. thompsonae</i>
				<i>W. ornata</i>
			middle <i>M. diversus</i>	<i>W. walpawanaensis</i>
			lower <i>M. diversus</i>	<i>W. hyperacantha</i>
Paleocene	upper <i>L. balmei</i>		<i>A. homomorpha</i>	
	lower <i>L. balmei</i>		<i>E. crassitabulata</i>	
				<i>T. evittii</i>
Late Cretaceous	Maastrichtian	<i>T. longus</i>	<i>M. druggii</i>	
	Campanian	<i>T. liliei</i>	<i>I. korojonense</i>	
		<i>N. senectus</i>	<i>X. australis</i>	
	Santonian	<i>T. pachyexinus</i>	<i>N. aceris</i>	
	Coniacian		<i>I. cretaceum</i>	
			<i>O. porifera</i>	
	Turonian	<i>C. triplex</i>	<i>C. striatoconus</i>	
Cenomanian	<i>A. distocarinatus</i>	<i>P. infusorioides</i>		
Early Cretaceous	Albian	Late	<i>P. pannosus</i>	
		Middle	upper <i>C. paradoxa</i>	
		Early	lower <i>C. paradoxa</i>	
	Aptian		<i>C. striatus</i>	
			upper <i>C. hughesi</i>	
		lower <i>C. hughesi</i>		
	Barremian		<i>F. wonthaggiensis</i>	
	Hauterivian			
	Valanginian		upper <i>C. australiensis</i>	
	Berriasian		lower <i>C. australiensis</i>	
Juras.	Tithonian	<i>R. watheroensis</i>		

FIGURE 1

ZONATION FRAMEWORK

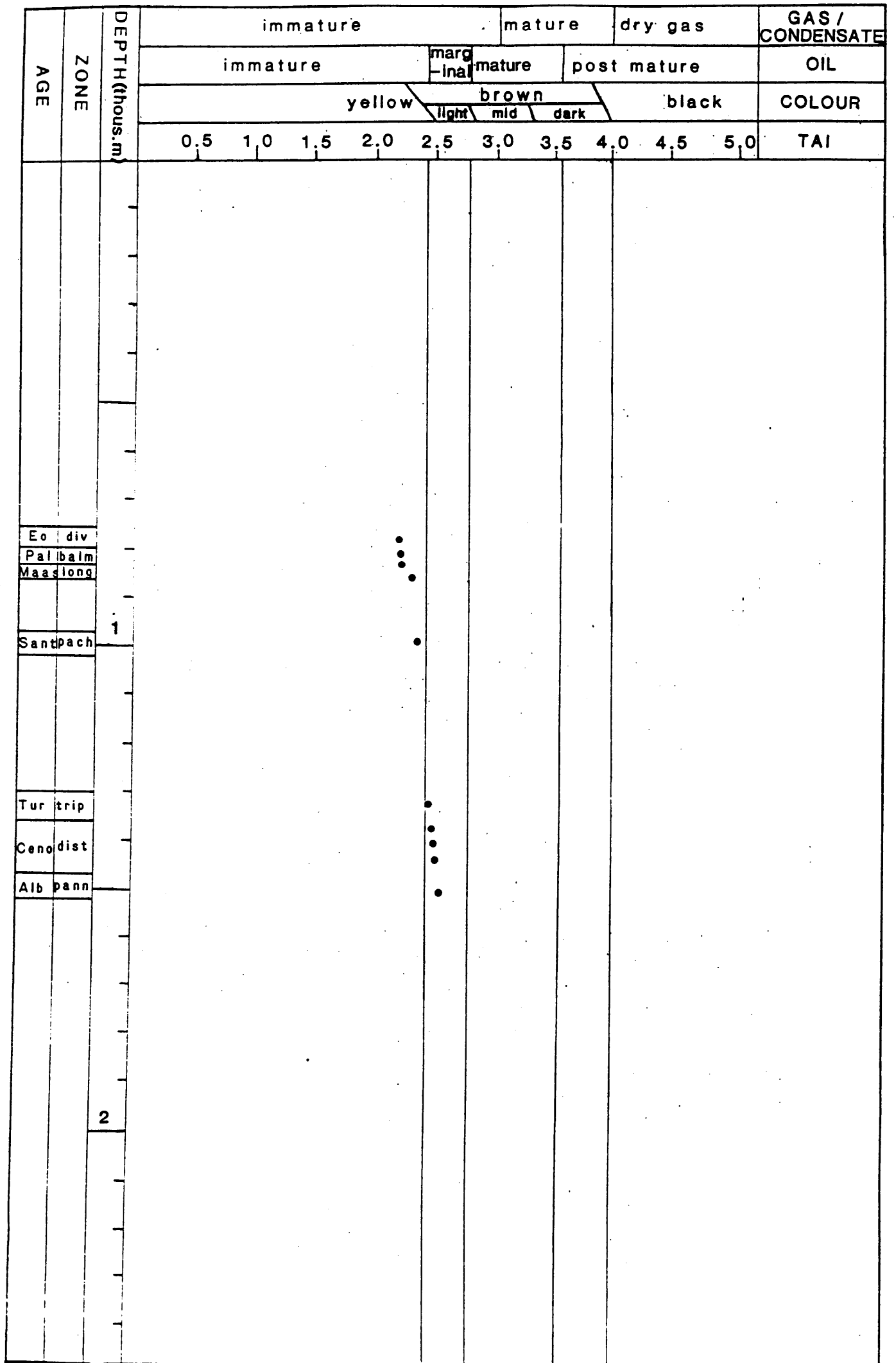


FIGURE 2 MATURITY PROFILE, SQUATTER-1

III PALYNOSTRATIGRAPHY

A. 790m (cutts) : M. diversus Zone (possibly upper)

Assignment to the Malvacipollis diversus Zone is clearly indicated at the top by the absence of younger indicators such as Proteacidites asperopolus, Nothofagidites falcatus etc, and at the base by oldest Cupaneidites orthoteichus, Intratriporopollenites notabilis, Spinozinocolpites prominatus and Malvacipollis diversus without older indicators. Subzonal assignment is problematic, as the subzones are defined on oldest occurrences which are easily caved in cuttings samples. Oldest Proteacidites clarus and P. kopiensis suggest middle M. diversus Zone or younger, and oldest Proteacidites pachypolus suggests upper M. diversus Zone or younger, but these taxa could all be caved from above. If all the taxa seen are in place, an upper M. diversus age would be indicated. Dilwynites and Proteacidites are dominant. Minor Cretaceous and Permian reworking were seen.

Dinoflagellates include frequent Muratodinium fimbriatum and rare Hafniasphera septata, consistent with the M. diversus assignment, but not sufficient to indicate a subzone.

Very nearshore marine environments are suggested by the low dinoflagellate content (10% of palynomorphs) and their low diversity (5 species). Pollen and spores are dominant and diverse.

These features are normally seen in the Pember Member of the Dilwyn Formation.

Light yellow spore colours indicate immaturity for hydrocarbon generation.

B. 810m (cutts) : L. balmei Zone

Assignment to the Lygistepollenites balmei Zone is indicated

at the top by youngest Gambierina edwardsii, G. rudata and L. balmei, and at the base by oldest L. balmei without older indicators. Proteacidites grandis and P. incurvatus are present and suggest the upper subzone, but they could be caved. Tetracolporites verrucosus is also present and suggests the lower subzone. However, only a single specimen was seen and so confidence is low (it could be reworked). Minor Cretaceous and Permian reworking were seen. Dilwynites, Falcisporites and Cyathidites are frequent.

Dinoflagellates are dominated by Deflandrea speciosa, suggesting a general Paleocene age. Other taxa include Isabelidinium bakeri (suggesting the lower L. balmei Zone), and several obviously or probably caved taxa (Apectodinium hyperacantha, Deflandrea obliquipes, Wetzeliella articulata, Hafniasphaera septata and Muratodinium fimbriatum) and some obviously reworked taxa (Isabelidinium pellucidum). An unusual new reticulate Senoniasphaera was seen.

Nearshore marine environments are indicated by the moderate dinoflagellate content (20%) and moderate diversity (although some of the diversity is caved).

These features are normally seen in the Pebble Point Formation.

Light yellow spore colours indicate immaturity for hydrocarbons.

- C. 820m (cutts) : mixed assemblage presumed L. balmei with latest Cretaceous T. longus reworking.

Assignment of this sample is problematic. The majority of the assemblage is consistent with an L. balmei assignment (including dominant Haloragacidites harrisii with scarce Gambierina rudata and Jaxtacolpus peirensis. However, single specimens of Tricolpites sabulosus and Triporopollenites sectilis and six specimens of dinoflagellates suggest the latest Cretaceous T. longus Zone. Since late Cretaceous

specimens are rare, and many markers are missing, a Paleocene L. balmei Zone assignment is considered likely, with minor Late Cretaceous reworking. However, it is not impossible that the Cretaceous has been penetrated near the base of the cuttings interval. Obvious Eocene caving comprises about 5% of the assemblage.

Dinoflagellates are very scarce and either long ranging or obviously caved Eocene or presumably reworked Cretaceous (Isabelidinium coronatum). Only D. speciosus, H. tubiferum and G. retiintexta may be in place, suggesting a general Paleocene age.

Marginally marine environments are indicated by the very scarce low diversity "in place" dinoflagellate assemblage, and the diverse and common pollen and spores.

The L. balmei Zone assignment is normally seen in the Pebble Point Formation, while a T. longus assignment is normally seen in the Timboon/Paaratte Formations.

Yellow spore colours indicate immaturity for hydrocarbons.

- D. 840m (cutts) : T. longus Zone (M. druggii dinoflagellate Zone)

This sample is assigned to the Tricolpites longus Zone at the top on youngest T. longus, T. confessus, T. waiparaensis and Triporopollenites sectilis. T. longus in particular is relatively frequent and the numerous late Cretaceous indicators leave no doubt, in contrast to the sample above. At the base, oldest T. longus and Tetracolporites verrucosus indicate the assignment. Proteacidites and Phyllocladidites mawsonii are common.

Dinoflagellates include Manumiella coronata, clearly indicating assignment to the M. druggii Dinoflagellate Zone. Other significant taxa include Isabelidinium pellucidum and some specimens of I. pellucidum showing affinities towards I.

korojonense.

Marginal marine environments are indicated by the very rare (1%) of very low diversity (3 species) of dinoflagellates.

These features are normally seen in the Timboon/Paaratte interval.

Spore colours of yellow indicate immaturity for hydrocarbon generation.

- E. 1000m (cutts) : T. pachyexinus Zone (N. aceras Dinoflagellate Zone)

Assignment is indicated at the top by the absence of younger indicators such as Nothofagidites senectus, and at the base by oldest Tricolporites pachyexinus. The absence of Amosopollis cruciformis suggests the upper part of the zone, and is consistent with the dinoflagellate evidence. Proteacidites sp. dominate the samples, with frequent P. mawsonii and persistent Australopollis obscurus. Obvious Eocene caving comprises about 5% of palynomorphs.

Dinoflagellates include Nelsoniella aceras without Xenikoon australis and so indicate the N. aceras Dinoflagellate Zone, confirming the spore-pollen assignment. Heterosphaeridium heteracanthum and Trithyrodinium spp. dominate.

Nearshore to marginal marine environments are indicated by the dinoflagellate content (10% of palynomorphs) and their very low diversity (3 species).

These features are normally seen in the Paaratte Formation.

Yellow to yellow/light brown spore colours indicate immaturity for hydrocarbon generation.

- F. 1310m (cutts) : C. triplex Zone

Assignment to the Clavifera triplex Zone is indicated at the top on youngest Appendicisporites distocarinatus and at the base on oldest Clavifera triplex and P. mawsonii considered to be in place. Younger indicators include Nothofagidites senectus (suggesting the N. senectus or younger zones) and Ornamentifera sentosa (suggesting the T. pachyexinus or younger zones), but their light spore colours and the other evidence show that they are caved. Eocene caving comprises about 5% of the assemblage, but inertinite dominates the sample. Gleicheniidites is common.

Dinoflagellates are not age diagnostic and are partly caved from younger horizons.

Nearshore marine environments are indicated by the low dinoflagellate content (10%) and diversity (5 species).

These features are normally seen in the Belfast Mudstone and Flaxmans Formation.

Yellow/light brown spore colours indicate early marginal maturity for oil, and immaturity for gas/condensate.

- G. 1390m (cutts)-1420m (cutts) : A. distocarinatus Zone (P. infusorioides Dinoflagellate Zone)

Assignment to the Appendicisporites distocarinatus Zone is indicated at the top by the absence of younger indicators considered to be in place, a downhole influx of A. distocarinatus and A. tricornitatus, and the dinoflagellate evidence. C. triplex, A. obscurus and P. mawsonii in this interval show light spore colours, indicating their caved provenance. Gleicheniidites and Falcisporites are the most common forms. Eocene caving is generally rare, comprising 2-3% of palynomorphs. At the base of the interval, a downhole increase of spore diversity (including the typically Early Cretaceous forms Cicatricosisporites australiensis, Trilobosporites trioreticulosus and Triporoletes reticulatus) suggests proximity to shoreline.

Dinoflagellates include a distinct downhole influx of Cribooperidinium edwardsii at the interval top, indicating penetration of the Palaeohystrichophora infusorioides Dinoflagellate Zone. The C. edwardsii/Chlamydophorella nyei association is a useful local assemblage. Other taxa are either caved or long ranging.

Very nearshore marine environments are indicated by the very low dinoflagellate content (5% or less) and very low diversity (2-3 species considered in place).

These features are normally seen in the Flaxmans/Waare interval.

Light brown spore colours indicate marginal maturity for oil, and immaturity for gas/condensate.

H. 1500m (cutts) : P. pannosus Zone

Assignment to the Phimopollenites pannosus Zone is indicated at the top by youngest Coptospora paradoxa, which is coincident with a major palynofacies change from inertinite domination above, to liptinite/vitrinite below, and a downhole increase in diversity and content especially of taxa like Balmesporites holodictyus and Cicatricosisporites spp. Appendicisporites are notably absent. The Zone base is defined by oldest P. pannosus, although this could conceivably be caved from the Late Cretaceous, and this sample belong to the upper C. paradoxa Zone. In the absence of sidewall cores, these possibilities cannot be resolved. Cyathidites is dominant, with frequent Cicatricosisporites australiensis, Gleicheniidites and Microcachryidites antarcticus. Eocene caving comprises 3% and Late Cretaceous caving comprises 10% of palynomorphs.

Dinoflagellates are extremely rare and spore colours suggest that they are probably caved. Environments are therefore probably non-marine.

These features are normally seen in the topmost Eumeralla Formation.

Light brown spore colours indicate marginal maturity for oil, and immaturity for gas/condensate.

IV CONCLUSIONS

A. Geological

Given the log picks supplied, there appears to be no major problem. Top Eumeralla at 1425m is consistent, but major erosion of the Eumeralla cannot be demonstrated by palynology from the cuttings samples available. There is no obvious clean sand at the base of the Late Cretaceous, so no Waare Sandstone is identified, and the base Flaxmans Formation is therefore more subtle than usual. Top Flaxmans at 1352m, top Belfast at 1297m and top Paaratte/Timboon at 825m are generally compatible with the palynology.

The sample at 820m showing mixed latest Cretaceous and Tertiary suggests several possibilities. First, as discussed above, significant reworking of the Cretaceous into the basal Tertiary may have occurred. Second, the cuttings depths may not be exact against log depth, and the cuttings from 820m may include rock material below 825m. This seems unlikely, as the lithology below 825m appears to be clean sandstone from logs, and would probably be barren of palynomorphs. Third, the top Late Cretaceous may be picked low, and could lie as high as 812m (the palynology sample at 810m lacks Late Cretaceous), with a terminal Cretaceous shale being present between 812-825m, characterised by the spikey sonic response. Overall, the first possibility may be the most likely.

Top Pebble Point at 792 or 795m is consistent with the palynology, but as discussed above, the Pebble Point may comprise only the interval 795-812m (showing its typical high but relatively flat sonic response). The overlying Pember is also consistent, but in the absence of sidewall cores, the conformability or unconformability of the boundary cannot be determined. The existing data suggests that a sizable unconformity is possible.

B. Palynological

These data do not radically alter palynological concepts regarding the known sequence.

The Paleocene samples do however, contain some significant information which hints at possible detailed subdivision of the Pebble Point interval. The section studied herein is probably from the lower L. balmei Zone and appears to be dominated by Deflandrea speciosa types. The presence of Isabelidinium bakeri may also be a valid indicator of the lower part of the Pebble Point Formation. There appears to be scope for a project to erect a palynological subdivision of this interval, if drilling priorities warrant a more detailed understanding of the Pebble Point.

Eocene samples from recent wells also suggest that there is potential for a dinoflagellate zonation of the Pember/topmost Pebble Point based on acme horizons. For example, 790m contains frequent M. fimbriatum while 810m contains frequent Apectodinium spp. Such an acme based zonation could be worked easily in cuttings, and therefore overcome the problems of identifying the subdivisions of the M. diversus Zone (which are based on oldest occurrences).

C. Maturity

Spore Colours suggest marginal maturity at the well base, apparently in contrast to other data. Spore colour is a qualitative assessment made by eye. If other maturity data are instrumental and quantitative and therefore more repeatable, they would be favoured. However, those methods cannot distinguish between what is in place and what is caved in cuttings samples. A palynologist can determine what is in place and therefore assess the extent of caving, and account for it in his maturity evaluation.

V REFERENCES

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APPENDIX I

PALYNOMORPH OCCURRENCE DATA

0790.0 CUTTS
 0810.0 CUTTS
 0820.0 CUTTS
 0840.0 CUTTS
 1000.0 CUTTS
 1310.0 CUTTS
 1390.0 CUTTS
 1420.0 CUTTS
 1500.0 CUTTS

34	* APTEDIINIUM AUSTRALIENSE *	
35	* CORDOSPHAERIDIUM MULTISPINOSUM *	
36	* DEFLANDREA DILYNNENSIS *	
37	* DEFLANDREA MEDCALFII *	
38	* DEFLANDREA OBLIQUIPE *	
39	* DEFLANDREA SPECIOSUS *	
40	* DEFLANDREA STRIATA *	
41	* FIBROCYSTA BIPOLARE *	
42	* GLAPHYROCYSTA RETIINTEXTA *	
43	* HYSTRICHOSPHAERIDIUM TUBIFERUM *	
44	* ISABELIDIINIUM BAKERI *	
45	* ISABELIDIINIUM PELLUCIDUM *	
46	* OPERCULODINIUM CENTROCARPUM *	
47	* OPERCULODINIUM SP. *	
48	* SENONIASPHAERA SP. *	
49	* NETZELIELLA ARTICULATA *	
50	AMOSOPOLLIS CRUCIFORMIS	
51	DACRYCARPITES AUSTRALIENSIS	
52	DILYNNITES TUBERCULATUS	
53	FALCISPORITES SIMILIS	
54	GAMBIERINA EDWARDSII	
55	GAMBIERINA RUDATA	
56	HERKOSPORITES ELLIOTTII	
57	LATROBOSPORITES CRASSUS	
58	LYGISTEPOLLENITES BALMEI	
59	NOTHOFAGIODES BRACHYSPINULOSUS	
60	PEROMONOLITES VELLOSUM	
61	PHYLLOCLADITES MANSONII	
62	PHYLLOCLADITES RETICULOSACCATUS	
63	PHYLLOCLADITES VERRUCOSUS	
64	PODOSPORITES MICROSCCATUS	
65	PROTEACIODES PALISADUS	
66	PROTEACIODES TENUIEXIMUS	

0790.0 CUTTS
 0810.0 CUTTS
 0820.0 CUTTS
 0840.0 CUTTS
 1000.0 CUTTS
 1310.0 CUTTS
 1390.0 CUTTS
 1410.0 CUTTS
 1420.0 CUTTS
 1500.0 CUTTS

67 TETRACOLPORITES VERRUCOSUS
 68 VERRUCOSISPORITES KOPUKUENSIS
 69 * AREOLIGERA CORONATA *
 70 * CORDOSPHAERIDIUM INODES *
 71 * MANUMIELLA CORONATA *
 72 CERATOSPORITES EQUALIS
 73 JAMTACOLPUS PEIRATUS
 74 PROTEACIDITES TUBERCULIFORMIS
 75 STEREISPORITES REGIUM
 76 TRICOLPITES SABULOSUS
 77 TRIPOROPOLLENITES SECTILIS
 78 * NUMMUS MONOCULATUS *
 79 NOTHOFAGIDITES ENDURUS
 80 NOTHOFAGIDITES SENECTUS
 81 TRICOLPITES CONFESSUS
 82 TRICOLPITES LONGUS
 83 TRICOLPITES WAIPARAENSIS
 84 * HETEROSPHAERIDIUM *
 85 * NELSONIELLA ACERAS *
 86 * TRITHYROIDINIUM "RETICULATA" *
 87 BALMEISPORITES HOLODICTYUS
 88 CAMEROZOSPORITES OHAIENSIS
 89 CICATRICOSISPORITES AUSTRALIENSIS
 90 CICATRICOSISPORITES CUNEIFORMIS
 91 CICATRICOSISPORITES HUGHESI
 92 CICATRICOSISPORITES LUDBROOKIAE
 93 COROLLINA TOROSUS
 94 CRYBELOSPORITES STRIATUS
 95 OSMUNDACIDITES WELLMANNII
 96 PHIMOPOLLENITES PANNOSUS
 97 RETITRILETES CIRCOLUMENUS
 98 TRICOLPORITES PACHYXINUS
 99 TRIPOROLETES RETICULATUS

90.0 CUTTS
 0810.0 CUTTS
 0820.0 CUTTS
 0840.0 CUTTS
 1000.0 CUTTS
 1310.0 CUTTS
 1390.0 CUTTS
 1410.0 CUTTS
 1420.0 CUTTS
 1500.0 CUTTS

1100	* ODONTOCHITINA OPERCULATA *
1101	* OLIGOSPHAERIIDIUM COMPLEX *
1102	APPENDICISPORITES DISTOCARINATUS
1103	CINGUTRILETES CLAVUS
1104	CYATHIDITES AUSTRALIS
1105	CYATHIDITES MINOR
1106	CYCADOPITES FOLLICULARIS
1107	FORAMINISPORIS DAILYI
1108	MICROCACHRYIDITES ANTARCTICUS
1109	NEORAISTRICKIA TRUNCATA
1110	ORNAMENTIFERA SENTOSA
1111	PERINOPOLLENITES ELATOIDES
1112	* APTEA POLYMORPHA *
1113	* CHLAMYDOPHORELLA NYEI *
1114	* CLEISTOSPHAERIIDIUM SP. *
1115	* CRIBROPERIDIUM EDWARDSI *
1116	* CYCLONEPHELIUM COMPACTUM *
1117	* ISABELIDIINIUM BALMEI *
1118	* ISABELIDIINIUM COOKSONIAE *
1119	* SPINIFERITES RAMOSUS *
1120	ANNULISPORITES FOLLICULOSA
1121	APPENDICISPORITES TRICORNITATUS
1122	DICTYOTOSPORITES COMPLEX
1123	FORAMINISPORIS WINTHAGGIENSIS
1124	LYCOPODIACIDITES ASPERATUS
1125	PEROTRILETES JUBATUS
1126	PEROTRILETES SP.
1127	TRILETES TUBERCULIFORMIS
1128	ARAUCARIACITES AUSTRALIS
1129	CALLIALASPORITES DAMPIERI
1130	LEPTOLEPIDITES VERRUCATUS
1131	CONTIGNISPORITES COOKSONIAE
1132	CORONATISORA PERFORATA

0790.0	CUTTS	1133	ISCHYOSPORITES PUNCTATUS
0810.0	CUTTS	1134	KLUKISPORITES SCABERIS
0820.0	CUTTS	1135	TRILOBOSPORITES TRIORETICULOSUS
0840.0	CUTTS	1136	TRIPOROLETES RADIATUS
1000.0	CUTTS	1137	ANTULSPORITES VARIGRANULATUS
1310.0	CUTTS	1138	CAMEROZONOSPORITES BULLATUS
1390.0	CUTTS	1139	COPTOSPORA PARADOXA
1410.0	CUTTS	1140	FALCISPORITES GRANDIS
1420.0	CUTTS	1141	FORAMINISPORIS ASYMMETRICUS
1500.0	CUTTS	1142	LEPTOLEPIDITES MAJOR
		1143	RETTITRILETES FACETUS
		1144	TRILOBOSPORITES PURVERULENTUS
		1145	TRIPOROLETES SIMPLEX

SPECIES LOCATION INDEX

) Index numbers are the columns in which species appear.

INDEX NUMBER	SPECIES
31	* APECTODINIUM HOMOMORPHA (L.) *
32	* APECTODINIUM HOMOMORPHA (SH.) *
33	* APECTODINIUM HYPERCANTHA *
112	* APTEA POLYMORPHA *
34	* APTEODINIUM AUSTRALIENSE *
69	* AREOLIGERA CORONATA *
1	* AREOLIGERA SENONENSIS *
113	* CHLAMYDOPHORELLA NYEI *
114	* CLEISTOSPHAERIDIUM SP. *
70	* CORDOSPHAERIDIUM INODES *
35	* CORDOSPHAERIDIUM MULTISPINOSUM *
115	* CRIBROPERIDIUM EDWARDSI *
116	* CYCLONEPHELIUM COMPACTUM *
36	* DEFLANDREA DILWYNENSIS *
37	* DEFLANDREA MEDCALFII *
38	* DEFLANDREA OBLIQUIPES *
39	* DEFLANDREA SPECIOSUS *
40	* DEFLANDREA STRIATA *
41	* FIBROCYSTA BIFOLARE *
42	* GLAPHYROCYSTA RETIINTEXTA *
2	* HAFNIASPHAERA SEPTATA *
84	* HETEROSPHAERIDIUM *
43	* HYSTRICHOSPHAERIDIUM TUBIFERUM *
44	* ISABELIDINIUM BAKERI *
117	* ISABELIDINIUM BALMEI *
118	* ISABELIDINIUM COOKSONIAE *
45	* ISABELIDINIUM PELLUCIDUM *
71	* MANUMIELLA CORONATA *
3	* MURATODINIUM FIMBRIATUM *
85	* NELSONIELLA ACERAS *
78	* NUMMUS MONOCULATUS *
100	* ODONTOCHITINA OPERCULATA *
101	* OLIGOSPHAERIDIUM COMPLEX *
46	* OPERCULODINIUM CENTROCARPUM *
47	* OPERCULODINIUM SP. *
48	* SENONIASPHAERA SP. *
119	* SPINIFERITES RAMOSUS *
86	* TRITHYRODINIUM "RETICULATA" *
49	* WETZELIELLA ARTICULATA *
50	AMOSOPOLLIS CRUCIFORMIS
120	ANNULISPORITES FOLLICULOSA
137	ANTULSPORITES VARIGRANULATUS
) 102	APPENDICISPORITES DISTOCARINATUS

121 APPENDICISPORITES TRICORNITATUS
 128 ARAUCARIACITES AUSTRALIS
 4 AUSTRALOPOLLIS OBSCURUS
 87 BALMEISPORITES HOLODICTYUS
 129 CALLIALASPORITES DAMPIERI
 138 CAMEROZONOSPORITES BULLATUS
 88 CAMEROZONOSPORITES OHAIENSIS
 72 CERATOSPORITES EQUALIS
 89 CICATRICOSISPORITES AUSTRALIENSIS
 90 CICATRICOSISPORITES CUNEIFORMIS
 91 CICATRICOSISPORITES HUGHESI
 92 CICATRICOSISPORITES LUDBROOKIAE
 103 CINGUTRILETES CLAVUS
 5 CLAVIFERA TRIPLEX
 131 CONTIGNISPORITES COOKSONIAE
 139 COPTOSFORA PARADOXA
 93 COROLLINA TOROSUS
 132 CORONATISFORA PERFORATA
 94 CRYBELOSFORITES STRIATUS
 6 CUFANIEIDITES ORTHOTEICHUS
 104 CYATHIDITES AUSTRALIS
 105 CYATHIDITES MINOR
 7 CYATHIDITES SFLENDENS
 8 CYATHIDITES SPP.
 106 CYCADOPIITES FOLLICULARIS
 51 DACRYCARPITES AUSTRALIENSIS
 122 DICTYOTOSPORITES COMPLEX
 9 DILWYNITES GRANULATUS
 52 DILWYNITES TUBERCULATUS
 10 ERICIPITES SCABRATUS
 140 FALCISPORITES GRANDIS
 53 FALCISPORITES SIMILIS
 141 FORAMINISPORIS ASYMMETRICUS
 107 FORAMINISPORIS DAILYI
 123 FORAMINISPORIS WONTHAGGIENSIS
 54 GAMBIERINA EDWARDSII
 55 GAMBIERINA RUDATA
 11 GLEICHENIIDITES
 12 HALORAGACIDITES HARRISII
 56 HERKOSPORITES ELLIOTTII
 13 INTRATRIFOROPOLLENITES NOTABILIS
 14 ISCHYOSPORITES GREMIUS
 133 ISCHYOSPORITES FUNCTATUS
 73 JAXTACOLFUS FEIRATUS
 134 KLUKISPORITES SCABERIS
 57 LATROBOSFORITES CRASSUS
 15 LATROBOSFORITES OHAIENSIS
 142 LEPTOLEPIDITES MAJOR
 130 LEPTOLEPIDITES VERRUCATUS
 124 LYCOPODIACIDITES ASFERATUS

58 LYGISTEPOLLENITES BALMEI
 16 LYGISTEPOLLENITES FLORINII
 17 MALVACIFOLLIS DIVERSUS
 18 MALVACIFOLLIS SUBTILIS
 108 MICROCACHRYIDITES ANTARCTICUS
 109 NEORAISTRICKIA TRUNCATA
 59 NOTHOFAGIDITES BRACHYSPINULOSUS
 79 NOTHOFAGIDITES ENDURUS
 80 NOTHOFAGIDITES SENECTUS
 110 ORNAMENTIFERA SENTOSA
 95 OSMUNDACIDITES WELLMANII
 111 PERINOPOLLENITES ELATOIDES
 19 PERIPOROPOLLENITES POLYORATUS
 60 PEROMONOLITES VELLOSUM
 125 PEROTRILETES JUBATUS
 126 PEROTRILETES SP.
 96 PHIMOPOLLENITES FANNOSUS
 61 PHYLLOCLADIDITES MAWSONII
 62 PHYLLOCLADIDITES RETICULOSACCATUS
 63 PHYLLOCLADIDITES VERRUCOSUS
 64 PODOSPORITES MICROSACCATUS
 20 PROTEACIDITES ANNULARIS
 21 PROTEACIDITES CLARUS
 22 PROTEACIDITES GRANDIS
 23 PROTEACIDITES INCURVATUS
 24 PROTEACIDITES KOPIENSIS
 25 PROTEACIDITES PACHYPOLUS
 65 PROTEACIDITES PALISADUS
 26 PROTEACIDITES SPP.
 66 PROTEACIDITES TENUIEXINUS
 74 PROTEACIDITES TUBERCULIFORMIS
 27 RETITRILETES AUSTROCLAVATIDITES
 97 RETITRILETES CIRCOLUMENUS
 143 RETITRILETES FACETUS
 28 SPINIZONOCOLPITES FROMINATUS
 29 STEREISPORITES (TRIPUNCTISPORIS) SPP.
 30 STEREISPORITES ANTIQUASPORITES
 75 STEREISPORITES REGIUM
 67 TETRACOLPORITES VERRUCOSUS
 81 TRICOLPITES CONFESSUS
 82 TRICOLPITES LONGUS
 76 TRICOLPITES SABULOSUS
 83 TRICOLPITES WAIPARAENSIS
 98 TRICOLPORITES PACHYEXINUS
 127 TRILETES TUBERCULIFORMIS
 144 TRILOBOSPORITES FURVERULENTUS
 135 TRILOBOSPORITES TRIORETICULOSUS
 136 TRIFOROLETES RADIATUS
 99 TRIPOROLETES RETICULATUS
 145 TRIPOROLETES SIMPLEX
 77 TRIPOROPOLLENITES SECTILIS
 68 VERRUCOSISPORITES KOPUKUENSIS